



April 27, 2018

Linda Culpepper

Director, Division of Water Resource, North Carolina Department of Environmental Quality 217 West Jones Street 1611 Mail Service Center Raleigh, NC 27699

Re: IMAC Development Petition

Dear Ms. Culpepper,

Pursuant to 15A NCAC 02L .0202(c), Chemours is petitioning for the establishment of an interim maximum allowable concentration for 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)propanoate (CAS # 13252-13-6, also referred to as hexafluoropropylene oxide dimer acid [HFPO-DA] and known by the trade name GenX). Attached you will find the petition that includes relevant toxicological and epidemiological data, study results, and calculations necessary to establish a standard in accordance with 15A NCAC 02L .0202(d).

Thank you for considering this petition. If you have any questions or need additional information, please feel free to contact me at 704-560-6435.

Respectfully submitted,

Kin Haron

Kevin Garon

Project Director, Chemours Corporate Remediation Group

Kevin.Garon@Chemours.com

CC: Michael E. Scott

Director, Division of Waste Management, North Carolina Department of Environmental Quality 217 West Jones Street 1646 Mail Service Center Raleigh, NC 27699



To:

Kevin Garon – Chemours CRG Project Director

CC:

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Project name:

Fayetteville Regulatory Support

Project ref: 60571508

From:

Dana McCue, AECOM Principal Risk Assessor

Date:

April 27, 2018

Memorandum

Subject: IMAC Development for 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)propionic acid (CAS # 13252-13-6) - Chemours Fayetteville Works, Fayetteville, North Carolina

AECOM has prepared this memorandum on behalf of The Chemours Company FC, LLC (Chemours) for the Fayetteville Works facility located near Duart Township in Bladen County, North Carolina (the site). The site was owned by E. I. du Pont de Nemours and Company (DuPont) until July 2015.

The purpose of this memorandum is to describe the derivation of an Interim Maximum Allowable Concentration (IMAC) for 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)propanoate (CAS # 13252-13-6, also referred to as hexafluoropropylene oxide dimer acid [HFPO-DA] and known by the trade name GenX). A review of the available literature indicates that a suitable body of toxicity information is available to support the derivation of toxicity reference values and the IMAC. The derivation was undertaken because HFPO-DA does not have a promulgated groundwater standard under Title 15A of the North Carolina Administrative Code, Subchapter 02L (15A NCAC 02L).

Section 0202(d) under 15A NCAC 02L establishes the IMAC as the lesser of the following six criteria:

- Systemic threshold concentration;
- 2. Concentration which corresponds to an incremental lifetime cancer risk of 1 x 10⁻⁶;
- 3. Taste threshold limit value;
- Odor threshold limit value;
- 5. Maximum contaminant level (MCL); or
- National secondary drinking water standard.

Based on a 2-year chronic oral gavage study of HFPO-DA in rats, an oral reference dose (RfD) of 0.01 milligrams per kilogram per day (mg/kg/day) was calculated that served as the most appropriate and relevant point of departure (POD) for establishing a systemic threshold concentration for HFPO-DA (70 micrograms per liter [μ g/L]). HFPO-DA is not mutagenic or genotoxic, and is not expected to exhibit a carcinogenic response in mammalian tissues. No odor or taste threshold data is available for HFPO-DA. There is no assertion of any taste or odor issues related to HFPO-DA. In addition, HFPO-DA does not have a MCL or secondary drinking water standard. As a result, the systemic threshold

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concentration is the basis for the proposed IMAC value (70 μ g/L). Technical information necessary to establish an IMAC for HFPO-DA consistent with 15A NCAC 02L is detailed herein.

General Information

HFPO-DA is a member of a family of chemical compounds know as per- and polyfluoroalkyl substances (PFAS). PFAS encompass a wide universe of substances with very different physical and chemical properties, including gases (for example, perfluorobutane), liquids (for example, fluorotelomer alcohols), surfactants (for example, perfluorooctane sulfonate), and solid material high-molecular weight polymers (for example, polytetrafluoroethylene [PTFE]) (Interstate Technology Regulatory Council [ITRC], 2017).

Perfluoroalkyl acids (PFAAs) are some of the most basic PFAS molecules. They are essentially non-degradable and currently are the class of PFAS most commonly tested for in the environment. Concern regarding the persistence, bioaccumulation, and possible ecological and human health effects of long-chain PFAAs has led manufacturers to develop replacement short-chain PFAS chemistries that should not degrade to longchain PFAAs (United States Environmental Protection Agency [USEPA], 2006; OECD, 2017). Examples of this trend are replacement PFAS that have been developed for use as processing aids in the manufacturing of fluoropolymers (ITRC, 2017 and Rae et al., 2015). One of these processing aids is HFPO-DA.

HFPO-DA is the precursor to the ammonium salt of 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoic acid (CAS #62037-80-3) and is the environmentally relevant form in water if released to the environment. Chemical and physical properties for HFPO-DA and HFPO-DA ammonium salt include the following:

Analyte	•	Ammonium Salt 62037-80-3)	HFPO-DA (CAS #13252-13-6)		
Property	Value	Source	Value	Source	
Molecular Weight (g/mol)	347	Rae et al., 2015	330	EPA CompTox Database (2018)	
Solubility (mg/L)	Infinitely soluble in water	Nixon and Lezotte, 2008a	Infinitely soluble in water	Nixon and Lezotte, 2008b	
Henry's Law Constant (atm-m ³ /mol)	2.37 x 10 ⁻¹⁰	EPA CompTox Database (2018)	2.37 x 10 ⁻¹⁰	EPA CompTox Database (2018)	
Vapor Pressure (Pa) at 20 deg C	0.01 (solid)	Rae et al., 2015	306 (liquid)	Nixon and Lezotte, 2008b	
Soil Adsorption Coefficient (Koc) (L/Kg)	12	Bloxham, 2008	Not Available	'	
Bioconcentration Factor (BCF)	6.27	EPA CompTox Database (2018)	6.27	EPA CompTox Database (2018)	

Systemic Threshold Concentration

A systemic threshold concentration for HFPO-DA can be calculated using the following equation and exposure assumptions specified in 15A NCAC 02L .0202(d) and North Carolina's *Division of Water Quality Standard Operating Procedure (SOP) for Reviewing Groundwater Standards Established Pursuant to 15A NCAC 02L.0200*:

RfD x BW x RSC DWI

where:

RfD = Reference dose, oral (chemical-specific, mg/kg/day)

BW = Body Weight (70 kg)

RSC = Relative source contribution (0.2 for organics)

DWI = Drinking water intake (2 liters/day)

The RfD is a pathway-specific (e.g., oral) estimate of a daily chemical intake per unit body weight that is likely to be without deleterious effects (chronic) for a lifetime of exposure, including sensitive subpopulations (USEPA, 1989). RfDs are derived from experimental data and include safety factors to account for differences among species and within populations and other uncertainties in the experimental data.

Consistent with 15A NCAC 2L.0202(e), in the absence of toxicity values available from USEPA, California EPA or Agency for Toxic Substances and Disease Registry (ATSDR) sources, other relevant, published health risk assessment data, and scientifically valid peer-reviewed published toxicological data were reviewed and evaluated to develop an oral reference dose. **Attachment A** to this memorandum details the review of the available toxicological studies. **Table 1** provides a summary of the relevant study findings.

As detailed in **Attachment A** and summarized in **Table 1**, seven toxicological studies were identified as relevant to the development of the IMAC for HFPO-DA. All available toxicological studies were performed using the HFPO-DA ammonium salt. As discussed in Attachment A, upon dissolution in the gut and absorption and distribution of the acid and the salt in the test organism, the dissociated acid form would be present and thus a read-across (i.e., toxicological equivalency) of the salt to the acid form is justified for an evaluation of systemic effects and is consistent with other analyses of these data (Beekman, 2016).

The toxicological studies include four oral toxicity studies in rats (28-day subacute, 90-day subchronic, 2-year chronic and carcinogenicity, and a prenatal developmental study) and three oral toxicity studies in mice (28-day subacute study, a 90-day subchronic study, and a 70+ day developmental and reproductive toxicity (DART) study). All seven of these studies followed USEPA Toxic Substances Control Act (TSCA) (40 CFR Part 792) Good Laboratory Practice (GLP) Standards, which are compatible with current Organization for Economic Cooperation and Development (OECD) GLP Standards. The GLP Standards require routine inspections of the study while in progress, and periodic audits and peer reviews by an independent auditing team. In addition, these studies were accepted for use by the European Chemicals Agency (ECHA) to assess the toxicity of HFPO-DA to humans and deemed reliable. The ECHA identified only the 2-year chronic study as the key study to evaluate chronic oral toxicity, with the other six studies being adequate as supporting studies.

The 2-year chronic study was also chosen as the relevant study to derive the RfD for use in the systemic threshold concentration calculation for the IMAC. The 2-year chronic and carcinogenicity rat study, performed by Rae, et al (2015), not only represents the most relevant exposure route for the development of an IMAC, but also is of sufficient duration and magnitude to permit the development of a reference dose (USEPA, 1989). Current USEPA Health Effects Test Guidelines (OPPTS 870.4300) were followed. Including 80 rats per sex per dose group, plus controls, the study is sufficiently robust AECOM

in experimental design to permit drawing statistically valid conclusions regarding the toxicity of HFPO-DA. There is no scientific justification to use the subchronic or subacute studies rather than the 2-year chronic study, especially when the shorter duration studies do not reveal any different types of toxicity or modes of action that might require additional consideration.

The general equation for developing an RfD is from USEPA's *Risk Assessment Guidance for Superfund* (USEPA, 1989):

RfD (mg/kg/day) = NOAEL/(UF1 x UF2 x UF3)

Where:

NOAEL = the no observed adverse effect level in animals (mg/kg/day)

UF1 = an uncertainty factor applied when extrapolating from animals to humans

UF2 = an uncertainty factor to account for variation in the general population, intended to protect sensitive subpopulations

UF3 = an uncertainty factor applied when deriving the RfD from a subchronic rather than a chronic study

Substituting the lowest (most conservative) no-observed-effect level from the Rae et al (2015) study into the equation yields an oral RfD value of 0.01 mg/kg/day. Rationale for the UF values used in the equation is detailed in **Attachment A**. Combining this oral RfD with the exposure assumptions specified in 15A NCAC 2L.0202(d) results in a systemic threshold concentration for HFPO-DA of 70 µg/L, which is equal to 70,000 nanograms per liter (ng/L). This value is similar in magnitude to the public health goal (PHG) of 71,000 ng/L[§] initially derived by the North Carolina Department of Health and Human Services (NC DHHS) in June 2017.

In July 2017, NC DHHS revised the PHG downward to 140 ng/L. This revised PHG was based on a 10-fold lower NOAEL (0.1 mg/kg/day) identified from a 28-day subacute (subchronic) mouse study. Similar to the June 2017 PHG, values used for body weight and drinking water intake were based on bottle-fed infants³. These exposure assumptions are inconsistent with those specified in 15A NCAC 2L for IMAC development and with assumptions used by NC DHHS previously to derive the PHG for another PFAS (PFOA)³. Therefore, the revised PHG derivation is not considered an appropriate systemic threshold concentration for the IMAC.

Since the oral RfD used in the PHG derivation was based on a subacute/subchronic study rather than the more robust and more appropriate chronic study, an evaluation of a benchmark dose (BMD)-based RfD was conducted for comparison purposes. The BMD approach, which involves dose-response modeling to obtain BMD levels (BMDLs), i.e., dose levels corresponding to specific response levels near the low end of the observable range of the data, incorporates and conveys more information than the NOAEL and lowest observed adverse effect level (LOAEL) process traditionally used for noncancer health effects (USEPA, 2012).

As discussed in more detail in **Attachment A**, the 2-year chronic rat study data were not amenable to BMD modeling owing to the lack of observed effects at the lower doses and a wide range between the LOAEL (50 mg/kg/day) and NOAEL (1 mg/kg/day), resulting in a best-fit BMDL of 38.2 mg/kg/day (for liver centrilobular necrosis) which is much higher than the NOAEL Thus, the appropriate point of departure (POD) for the 2-year chronic study is the more conservative NOAEL of 1 mg/kg/day.

⁶ Based on an oral RfD of 0.01 mg/kg/day, 7.8 kg body weight, 1.1 L/day and RSC=1

Note that USEPA for the PFOA/PFOS health advisory goal used a DWI for pregnant and lactating women rather than a bottle-fed infant. Stating that" ...on the developing fetus resulting from exposures that occur during gestation and lactation. These developmental endpoints are the most protective for the population at large and are effects that can carry lifetime consequences for a less than lifetime exposure..."
DWI of 2L/day, BW of 70 kg and RSC of 0.2 was used by DHHS for the PFOA PHG.

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For the subchronic mouse studies, the data for male mice were combined from the 90-day subchronic and 70+ day DART studies because the exposure durations were similar, the mice were the same strain (Crl:CD1(ICR)) and single-cell necrosis in the liver was the common adverse endpoint reported by both studies. These data were amenable to BMD analysis and resulted in a best-fit BMDL of 0.23 mg/kg/day using the multistage model. Substituting the BMDL into the RfD equation yields an oral RfD value of 0.0023 mg/kg/day. Combining this oral RfD with the IMAC exposure assumptions results in a systemic threshold concentration for HFPO-DA of 16,000 ng/L.

A summary of the systemic threshold concentrations derived with the varying PODs is provided in the following table:

RfD (mg/kg/day)	BW (kg)	DWI (L/day)	RSC	STC (ng/L)
0.01	70	2	0.2	70,000
0.0023	70	2	0.2	16,000

In summary, the 2-yr chronic rat study is the most appropriate and relevant toxicity study for setting the POD for the oral RfD derivation and systemic threshold concentration calculation. However, in the absence of the chronic study data, setting the POD using the BMD method based on subchronic data from the mouse has a stronger scientific basis than setting the POD using the 0.1 mg/kg/day NOAEL from the subchronic/subacute mouse studies.

Concentration which corresponds to an incremental lifetime cancer risk of 1 x 10⁻⁶

In vitro and in vivo test results for the ammonium salt of HFPO-DA are indicative of a material that is not mutagenic or genotoxic. Although positive results were observed in one in vitro clastogenicity study with Chinese Hamster Ovary cells, negative results were observed for in vitro studies in bacterial cells (Salmonella typhimurium and E. coli) and mammalian cells (mouse lymphoma cells). In vivo studies yielded negative results for chromosome aberrations (mouse bone marrow), micronucleus (mouse bone marrow), and Unscheduled DNA Synthesis (rat liver cells) (a summary of the in vitro and in vivo studies is provided as **Table 2**).

Likewise, HFPO-DA is not expected to exhibit a carcinogenic response in mammalian tissues. As concluded in the Rae et al study (2015) and re-affirmed by ECHA, the induction of liver tumors in female rats at 500 mg/kg/day, and the equivocal increase in pancreatic acinar and testicular interstitial cell tumors in male rats at 50 mg/kg/day are likely not relevant to humans based on the following: most research indicates that induction of these specific tumors in rats by non-genotoxic peroxisome proliferators is not relevant to humans (Cunningham et al., 2010; Klaunig et al., 2012; Corton et al., 2018); the test material was determined to be non-genotoxic based on a battery of *in vivo* and *in vitro* genotoxicity studies; liver tumors were produced only in females and only at doses associated with marked hepatic and systemic toxicity (including lethality); and clear thresholds were established for all tumor types.

Taste Threshold Limit Value

A taste threshold value for HFPO-DA was not found in Young et al. (1996). Therefore, an in-depth search of the major electronic databases that may contain scientifically, peer-reviewed published data was conducted. This included searching OECD's eChem Portal; EPA's Chemical Safety for Sustainability CompTox Dashboard; ECHA; and, the National Library of Medicine's Toxline® Database. A taste threshold limit value was not found in the search.

Odor Threshold Limit Value

An odor threshold for HFPO-DA was not found in Young et al. (1996) or Amoore and Hautala (1983). Therefore, an in-depth search of the major electronic databases that may contain scientifically, peer-reviewed published data was conducted. This included searching OECD's eChem Portal; EPA's Chemical Safety for Sustainability CompTox Dashboard; ECHA; and, the National Library of Medicine's Toxline® Database. An odor threshold limit value was not found in the search.

Maximum Contaminant Level (MCL)

Currently, there is no MCL for HFPO-DA (USEPA, 2009).

National Secondary Drinking Water Standard (Secondary MCL)

Currently, there is no secondary MCL for HFPO-DA (USEPA, 2009).

INAC Recommendation

Based on a 2-year chronic feeding study of HFPO-DA in rats, an oral RfD of 0.01 mg/kg-day was calculated that served as the most appropriate and relevant POD for establishing a systemic threshold concentration for HFPO-DA (70 μ g/L). HFPO-DA is not mutagenic or genotoxic, and is not expected to exhibit a carcinogenic response in mammalian tissues. No odor or taste threshold data is available for HFPO-DA. There is no assertion of any taste or odor issues related to HFPO-DA. In addition, HFPO-DA does not have a MCL or secondary drinking water standard. As a result, the systemic threshold concentration is the basis for the proposed IMAC value (70 μ g/L).

References

Ammore and Haulata. 1983. *Odor as an Aid to Chemical Safety: Odor Thresholds Compared with Threshold Limit Values and Volatilities for 214 Industrial Chemicals in Air and Water Dilution*. Jour. Appl. Toxicol, 3, 272-290 (as cited in Toxicological Assessment of Cholorethane, Diphenyl and Phenyl Ether, S&ME, Inc., May, 1994).

Beekman. 2016. Evaluation of Substances Used in the GenX Technology, Chemours Dordrecht. RIVM Letter Report 2016-0174. The Netherlands National Institute for Public Health and the Environment.

Bloxham. 2008. DuPont Study 17568-1675: Estimation of the Adsorption Coefficient (Koc) of HFPO Dimer Acid Ammonium Salt on Soil and Sludge. Study Author: Peter A. Bloxham, Ph.D. Study Date: September 11, 2008.

Corton, J.C., Peters, J.M. and Klaunig, J.E. *The PPARα-dependent rodent liver tumor response is not relevant to humans: addressing misconceptions*. Arch Toxicol (2018) 92: 83. https://doi.org/10.1007/s00204-017-2094-7

Cunningham ML, Collins BJ, Hejtmancik MR, Herbert RA, Travlos GS, Vallant MK, and Stout MD. 2010. Effects of the PPARα agonist and widely used antihyperlipidemic drug gemfibrozil on hepatic toxicity and lipid metabolism. PPAR Research, Volume 2010, Article ID 681963.

Interstate Technology Regulatory Council (ITRC). 2018. ITRC Fact Sheet: Naming Conventions and Physical and Chemical Properties of Per- and Polyfluoroalkyl Substances (PFAS). Available on-line: https://pfas-1.itrcweb.org/wp-

content/uploads/2018/03/pfas fact sheet naming conventions 3 16 18.pdf

Klaunig, J.E., Hocevar, B.A. and Kamendulis, L.M. 2012. *Mode of action analysis of perfluorooctanoic acid (PFOA) tumorigenicity and human relevance*. Reprod.Toxicol. 33 (4) (2012) 410–418.

Nixon and Lezotte. 2008a. DuPont Study 24129: Determination of the Water Solubility and Vapor Pressure of H-28308. Study Authors: Willard B. Nixon, Ph.D. and Frank J. Lezotte, B.S. Study Date: March 27, 2008.

Nixon and Lezotte. 2008b. DuPont Study 24128: Determination of the Water Solubility and Vapor Pressure of H-28307. Study Authors: Willard B. Nixon, Ph.D. and Frank J. Lezotte, B.S. Study Date: March 27, 2008.

Organization for Economic Co-ooperation and Development (OECD). 2017. *OECD Portal on Per and Poly Fluorinated Chemicals*. Available from: http://www.oecd.org/chemicalsafety/portal-perfluorinated-chemicals/

Peters, J.M., Cheung C. and Gonzales, F.J. 2005. *Peroxisome-proliferator-activated receptor-alpha and liver cancer: where do we stand?* J. Mol. Med. 83 (2005)774–785.

Rae, J.M. Caverly, Craig, L., Slone, T., Frame, S., Buxton, L.W. and Kennedy, G.L. 2015. *Evaluation of chronic toxicity and carcinogenicity of ammonium 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoate in Sprague–Dawley rats*. Toxicology Reports 2 (2015) 939–949.

United States Environmental Protection Agency (USEPA). 1989. Risk Assessment Guidance for Superfund – Volume I, Human Health Evaluation Manual – Part A. EPA/540/1-89/002. December 1989.

USEPA. 2006. "2010/2015 PFOA Stewardship Program." *EPA-HQ-2003-0012-1071*. http://www.epa.gov/opptintr/pfoa/pubs/pfoastewardship.htm

USEPA. 2009. National Primary Drinking Water Regulations. EPA 816-F-09-004. Office of Groundwater and Drinking Water. May 2009. https://www.epa.gov/ground-water-and-drinking-water-national-primary-drinking-water-regulation-table

USEPA, 2012. Benchmark Dose Technical Guidance, EPA/100/R-12/001, June 2012.

Young, W.F., Horth, H., Crane, R., Ogden, T. and Arnott, M. 1996. *Taste and Odour Threshold Concentrations of Potential Potable Water Contaminants*. Water Research, 30:2, pp. 331-340.

Tables

TABLE 1 SUMMARY OF RELEVANT LITERATURE ON THE TOXICITY OF HFPO-DA (CAS # 13252-13-6)

IMAC Development for HFPO-DA Chemours Fayetteville Works, Fayetteville, North Carolina

Chudu Tibla	Study	Study	Study Guideline	Cassias	Dose (m	g/kg/day)	Ctudy Dun	.4:	Findings
Study Title	Date Author(s)		Study Guideline	Species	Male	Female	Study Duration		rinaings
Combined Chronic Toxicity/Oncogenicity Study 2-Year Oral Gavage Study in Rats (Rae et al, Toxicol. Rep. 2015)	3/28/2013	Craig, Lisa	OECD Guideline 453	Rat	0.1, 1, and 50	1, 50, and 500	up to 104 weeks in males and up to 101 weeks in females	Chronic	- NOAEL for chronic toxicity was 1 mg/kg/day in males and 50 mg/kg/day in females - The NOAEL in males is based on increases in focal cystic degeneration, focal necrosis, and centrilobular necrosis of the liver, with associated increases in cytotoxic liver enzymes, and equivocal increases in pancreatic acinar cell tumors and testicular interstitial (Leydig) cell tumors.
A 90-Day Oral (Gavage) Toxicity Study of H-28548 in Rats with a 28-Day Recovery	10/5/2009	Haas, Matthew C.	OECD Guideline 408	Rat	0.1, 10 and 100	10, 100 and 1000	90 days	Subchronic	- NOAEL was considered to be 10 mg/kg/day for males and 100 mg/kg/day for females based on evidence of regenerative anemia in males at 100 mg/kg/day and in females at 1000 mg/kg/day, as well as decreased survival in the 1000 mg/kg/day group females.
A 28-Day Oral (Gavage) Toxicity Study of H-28397 in Rats with a 28-Day Recovery	8/22/2008	Haas, Matthew C.	OECD Guideline 407	Rat	0.3, 3 and 30	3, 30 and 300	28 days	Subacute	- NOAEL was 30 mg/kg/day in males and 300 mg/kg/day in females - No effects on survival were noted.
An Oral (Gavage) Prenatal Developmental Toxicity Study of H-28548 in Rats	7/2/2010	Edwards, Tammye L.	OPPTS Guideline 870.3700 OECD Guideline 414	Rat		10, 100, and 1000	Gestation Day 6-20		- The NOAEL for maternal and developmental toxicity was considered to be 10 mg/kg/day based on mortality and lower mean body weight gains and food consumption at 1000 mg/kg/day and early deliveries, microscopic findings in the liver (focal necrosis), and lower mean fetal weights at 100 and 1000 mg/kg/day.
Subchronic Toxicity 90-Day Gavage Study in Mice	2/19/2010	Mackenzie, Susan A.	OECD Guideline 408	Mice	0.1, 0.5 and 5	0.1, 0.5 and 5	90 days	Subchronic	- NOAEL was 0.5 mg/kg/day in males and females, based on changes in clinical chemistry and histopathology indicative of liver toxicity in animals dosed with 5 mg/kg/day
An Oral (Gavage) Reproduction/Developmental Toxicity Screening Study of H-28548 in Mice	12/29/2010	Edwards, Tammye L.	OECD Guideline 421	Mice	0.1, 0.5 and 5	0.1, 0.5 and 5	70 days +		- There were no effects on reproduction (mating, fertility, or copulation indices, number of days between pairing and coitus, and gestation length). - The NOAEL for reproductive toxicity was 5 mg/kg/day, as no effects on reproduction were observed at any of the doses levels tested. - The NOAEL for systemic toxicity in male mice was 0.1 mg/kg/day based on adverse liver effects.
A 28-Day Oral (Gavage) Toxicity Study of H-28397 in Mice with a 28-Day Recovery	8/29/2008	Haas, Matthew C.	OECD Guideline 407	Mice	0.1, 3 and 30	0.1, 3 and 30	28 days	Subacute	- NOAEL was 0.1 mg/kg/day for males and 3 mg/kg/day for females Adverse effects were noted in the livers of males at 3 mg/kg/day and above and females at 30 mg/kg/day. These consisted of single cell necrosis of hepatocytes and correlative increases in liver enzymes. These effects were also reversible following the 4-week recovery period.

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TABLE 2 SUMMARY OF RELEVANT LITERATURE ON THE GENECTIC TOXICITY OF HFPO-DA (CAS # 13252-13-6)

IMAC Development for HFPO-DA
Chemours Fayetteville Works, Fayetteville, North Carolina

Study Type	Study Title	Study Date	Study Author(s)	Study Guideline	Type of Assay	Species/Strain	Result
In Vitro (Mutagenic effects - mammalian)	H-28548: In Vitro Mammalian Cell Gene Mutation Test (L5178Y/TK+/-Mouse Lymphoma Assay)	6/25/2008	Clarke, Jane J.	OECD Guideline 476	mammalian cell gene mutation assay	Mouse lymphoma L5178Y cells	Negative
In Vitro (Mutagenic effects - bacterial)	H-28072: Bacterial Reverse Mutation Test	8/13/2008	Donner, Maria E.	OECD Guideline 471	bacterial reverse mutation assay (Ames Study)	S. typhimurium TA 1535, TA 1537, TA 98 and TA 100; E. coli WP2 uvr A	Negative
In Vitro (Clastogenic effects - mammalian)	H-28072: In Vitro Mammalian Chromosome Aberration Test in Chinese Hamster Ovary Cells	9/23/2009	Glatt, Christine M.	OECD Guideline 473	in vitro mammalian chromosome aberration test	Chinese Hamster Ovary (CHO)	Positive
In Vivo (Clastogenic effects - mammalian)	H-28072: Unscheduled DNA Synthesis (UDS) Test with Mammalian Cells In Vivo	8/14/2007	Pant, Kamala; Sly, Jamie E.	OECD Guideline 486	unscheduled DNA synthesis	Rat/Sprague-Dawley	Negative
In Vivo (Clastogenic effects - mammalian)	H-28072: In Vivo Micronucleus and Chromosome Aberration Assay in Mouse Bone Marrow Cells	10/10/2007	Gudi, Ramadevi; Krsmanovic, Ljubica	OECD Guideline 474 and OECD Guideline 475	micronucleus assay and chromosome aberration assay	Mouse/ICR	Negative

Attachment A

Proposed Drinking Water Health Advisory Value for GenX: 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoic acid

Proposed Drinking Water Health Advisory Value for GenX: 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoic acid

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ABSTRACT

The widespread occurrence and subsequent environmental concerns of long-chain perfluoroalkyl acids (PFAAs) such as perfluorooctane sulfonate (PFOS) and perfluorooctanoic acid (PFOA) has led to the production and use of alternative polymer processing aids with more environmentally favorable chemical and biological properties. One alternative that has been used successfully is ammonium, 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoate, also known by its trade name GenX, and it has been detected in the environment near where it has been used in large quantities. This paper presents an analysis of the currently available toxicity data for GenX for the purpose of deriving a drinking water health advisory value using standard US EPA methodologies. A review of existing data on GenX revealed that a 2-year chronic study with rats provides the best available data on GenX toxicity for deriving a lifetime exposure health advisory. The No Observed Adverse Effect Level (NOAEL) for this study was 1.0 mg/kg/day and was used as the point of departure for deriving the health advisory. Using protective uncertainty factors of 10 for both interspecies and intraspecies and a conservative relative source contribution from water of 20%, a conservative lifetime health advisory value would be 70,000 ng/L for GenX. Furthermore, data on the elimination of GenX from mammals indicate it has a much faster elimination rate in humans compared to PFOA, PFOS and other legacy chemicals with an estimated half-life on the order of a few days at most.

INTRODUCTION

Per- and poly-fluorinated alkyl substances (PFAS) have been produced for over 70 years for use in industrial processes and manufacturing of consumer products (Kissa 2001; Buck 2011, Buck et al. 2015). The widespread use of PFAS has led to their release and detection in the environment (Heydebreck et al. 2015; Sun et al. 2016; Gebbink et al. 2017), most notably the long-chain perfluoroalkyl acids (PFAAs) such as perfluorooctane sulfonate (PFOS) and perfluorooctanoic acid (PFOA). Concern over the presence of these longer-chain PFAAs in the environment has led to development of alternative fluorinated substances with more environmentally favorable chemical and biological properties. An important class of these alternatives are the per- and polyfluorinated ether carboxylic acids (PFECAs) that have replaced ammonium perfluorooctanoate (APFO) as polymer processing aids (Feiring 1994; Buck 2015; Buck et al. 2011; Gordon 2011). Compared to the legacy long chain PFAAs, the shorter chain PFECAs have higher water solubility, much more rapid elimination from biological systems, are less toxic and expected to have very low accumulation in aquatic or terrestrial food webs (Ritter 2010; Buck et al. 2011; Caverly Rae et al. 2015; Gannon et al. 2016; Hoke et al. 2016).

A PFECA that has been used successfully as a polymer processing aid is ammonium, 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoate (EC 700-242-3, CAS 62037-80-3), also known by its trade name *GenX*, that is the conjugate base ammonium salt of the 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoic acid (Fig. 1, CAS 13252-13-6). This dimer acid is both the precursor to the salt in the production of GenX and the environmentally relevant form in water if GenX is released to the environment. The dimer acid form has been referred to as PFPrOPrA,

hexafluoropropylene oxide dimer acid, HFPO-DA, FRD 903 and as the trade name *GenX*. In this paper, the term *GenX* refers to the environmentally relevant deprotonated acid form of GenX. GenX has received significant attention recently owing to it being detected in the environment near where it has been used in large quantities: Cape Fear River, North Carolina, USA (Sun et al 2016); Parkersburg, West Virginia, USA; Dordrecht, Netherlands (Heydebreck et al. 2015; Gebbink et al. 2017); and Xiaoqing River, Shandong, China (Heydebreck et al. 2015). More specifically, some of the surface and groundwater where GenX has been detected serve as a drinking water source thus raising concern over possible adverse health effects associated with consumption of this water.

Figure 1. Chemical structure for 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoic acid.

GenX is not subject to proposed or promulgated drinking water regulations, nor is it on the US EPA Chemical Contaminants List (CCL) for unregulated chemicals, but the detection of GenX in North Carolina led to the issuance of a provisional lifetime health goal by the State of North Carolina (NC), USA. The NC provisional health goal was initially set at 71,000 ng/L, then revised to 140 ng/L, and now is subject to review by a Scientific Advisory Board (SAB) to the NC Department of Environmental Quality and NC Department of Health and Human Services, who may further revise the value based on the SAB recommendations (NCDEQ 2018).

This paper presents an analysis of the currently available toxicity data for GenX for the purpose of deriving a drinking water health advisory value (known as a health goal in the State of North Carolina). This paper is not intended to be as comprehensive as an US EPA Health Advisory document, but instead an initial summary and analysis of existing data to inform scientists contemplating further studies to assess exposure and toxicity, and for regulators and the public who may have concern about exposure to GenX and thus are making decisions regarding water use and/or treatment.

In this paper, all available data were treated to the standard methodologies used by the US EPA to derive lifetime drinking water health advisories and then recommend a proposed health advisory value for GenX based on the best available scientific data. This health advisory value is then compared to measured GenX exposures to provide perspective on the margin of exposure (or safety) at locations where GenX has been detected in drinking water sources.

NATURE OF THE CHEMICAL

The acid form of GenX is a liquid and the ammonium salt a white/colorless solid at ambient temperature (20 °C). Both are infinitely soluble in water above the pK_a (2.84) of the acid. The substance has a decomposition point of 150–160°C, a sublimation point of 130–140°C, a density of 1.7 g/cm³at 20°C, and a low vapor pressure of 0.01 Pa at 20°C (Caverly Rae et al. 2015; Gannon et al. 2016). Due to its high water solubility and negative charge at the pH of natural waters, GenX has a very low organic carbon-normalized adsorption coefficient (K_{oc}) to soil (log K_{oc} = 1.08) and sludge (log K_{oc} = 1.10) (Bloxham, 2008) and thus would not be expected to partition significantly onto environmental particles (e.g., soil, suspended or bedded sediment). This has implications for the fate of the chemical (e.g., it will not accumulate in sediments) and also for environmental sampling conducted to understand GenX fate or estimate exposure (e.g., focus should be on water and there should be little difference in measured values of GenX between whole water and filtered water samples).

In areas of large volume use of GenX, it has been found in soil, surface water, groundwater, and air (Heydebreck et al. 2015; Beekman 2016; Sun et al 2016; Gebbink et al. 2017; NCDEQ 2018), with very low concentrations in vegetables grown very near the GenX source (Mengelers 2018) and preliminary data indicating very low concentrations might be found in sediment (UNCW 2018).

Although detailed environmental fate data are not available at this time, GenX is expected to be relatively persistent in the environment and a biodegradability test indicated GenX was not readily biodegradable and not transformed structurally under the test conditions (Mitsubishi Chemical 2009). Given the high water solubility, GenX is expected to be mobile in the presence of water and thus would be expected to move from soil to groundwater and to surface waters in runoff or effluent discharge. The high water solubility and low volatility would cause any GenX emitted to air to return to the ground or surface water through wet deposition. This same high water solubility and very low K_{oc} would prevent GenX from partitioning strongly to soil or sediment or undergo significant bioaccumulation in aquatic food webs. Preliminary data with oysters exposed to high concentrations of GenX (100,000 ng/L) indicated very little accumulation in the oysters (UNCW 2018). The very high biological elimination rates (see below) would also prevent GenX from accumulating significantly in terrestrial food webs. Documents used by the European Chemicals Agency (ECHA) to review GenX support the above summary by concluding GenX "is unlikely to be a concern for aquatic bioaccumulation" (ECHA 2017a) and "is not a concern for terrestrial bioaccumulation" (ECHA 2017b).

HEALTH EFFECTS ASSESSMENT

The primary toxicological studies for GenX are four oral toxicity studies in rats (28-day subacute, 90-day subchronic, 2-year chronic and carcinogenicity, and a prenatal developmental study) and three oral toxicity studies in mice (28-day subacute study, a 90-day subchronic study, and 70+day developmental and reproductive toxicity (DART) study). This work is summarized in Table 1 (provided at the end of the document). All seven of these studies followed Good Laboratory

Practices (GLP). Five of these studies were reviewed by the ECHA to assess the toxicity of GenX to humans and all were deemed reliable. The ECHA identified only the 2-year chronic rat study (Craig 2013; Caverly Rae et al. 2015) as the Key Study, with the other studies being adequate as supporting studies only. The ECHA also included two 7-day non-GLP studies that are summarized below but not included in Table 1.

Additional GLP studies were conducted to evaluate acute toxicity, biodistribution/elimination, and genotoxicity. The oral LD50 (rat) was 1,750 mg/kg, the dermal LD50 was >5,000 mg/kg, the inhalation 4-h acute lethal concentration (ALC) was >3.6 mg/L vapor (Buck 2015). Genotoxicity was determined to be negative based on negative results in the following tests: Ames and chromosome aberration *in vitro* studies; and mouse bone marrow, mouse micronucleus, and rat unscheduled DNA synthesis (UDS) *in vivo* studies (Buck 2015).

Some non-GLP work has been published on GenX toxicity by academic researchers. *In vitro* responses (cytotoxicity and protein binding capacity) to GenX with a human liver cell line were investigated, but these data cannot be used directly for deriving a health advisory (Sheng et al. 2018). Immunomodulatory responses (T cell-dependent antibodies) to GenX exposure also were reported, but possible adverse effects were not found until very high doses (100 mg/kg) indicating that GenX is a very weak suppressor of immune function and thus would not be a factor in setting a health advisory value (Rushing et al. 2017).

Two non-GLP 7-day organ toxicity studies were conducted in mice at a single dose of 30 mg/kg/day that were identical except one used the GenX dimer acid (Nabb 2008a) and the other used the GenX dimer salt (Nabb 2008b, note this is also ECHA Study 006). The results were nearly identical: microscopic changes were limited to the liver and the study author indicated it was uncertain if these changes were related to the test substance. The value of these two studies is they demonstrate that the dimer acid and dimer salt are likely to have identical toxicities and thus a read-across (i.e., toxicological equivalency) of the salt to the acid form is justified and is consistent with other analyses of these data (Beekman 2016). This also is consistent with expected behavior, where both the acid and salt would be in the dissociated acid form following absorption and distribution in the test organism.

In the Key Study identified by ECHA, the 2-year chronic and carcinogenicity rat study (Craig 2013; Caverly Rae et al. 2015), the NOAELs were 1 mg/kg/day in males and 50 mg/kg/day in females; with the (more sensitive) NOAEL in the male rat based on observed effects at 50 mg/kg/day. These observed effects are manifested via a rodent-specific response to a PPARα-agonist exposure and thus is very likely a non-human-relevant mode of action (see below). These effects included focal cystic degeneration and necrosis, that study authors described as minimal to mild. These effects were observed only at 50 mg/kg/day (the Lowest Observed Adverse Effect level, LOAEL) in 5/70 animals and this response was not statistically significant. The shorter-term subchronic and subacute rat toxicity studies also reported liver effects that were considered adaptive and reversible responses to repeated chemical exposure that are commonly observed in rodents. In the developmental rat study, maternal toxicity was reported at this high dose. Of the liver at 100 mg/kg/day, and developmental effects were also reported at this high dose. Of

these four studies, the 2-year chronic study resulted in the lowest NOAEL for rats (1 mg/kg/day). Note that this study had no doses between 1 and 50 mg/kg/day (Table 1).

All of the mouse studies were considered by ECHA to be supporting studies only. They were only subchronic or subacute in duration, and they did not reveal any different types of toxicity or modes of action compared to the chronic rat study that might require additional consideration. Mice also have been shown to be less similar to primates than the rat with regard to GenX toxicokinetic behavior (see below).

The Appropriate Toxicity Study Duration for Lifetime Exposure Analysis

The US EPA risk assessment process uses the 1- or 2-year chronic study as the standard for assessing potential adverse health effects for a lifetime exposure duration and allows for subchronic studies to be used "when a chronic study is not available" (US EPA 2014a). For GenX, there is a complete 2-year chronic and carcinogenicity rat study that meets all the criteria established by the US EPA for use in a lifetime exposure analysis and was identified by ECHA as the Key Study (Craig 2013). There is therefore no scientific justification to use the subchronic or subacute studies as a surrogate for the 2-year chronic study, especially when the shorter duration studies do not reveal any different types of toxicity or modes of action that might require additional consideration. Thus, the 2-year chronic study was chosen as the Key Study for setting the proposed health advisory.

Identification of Adverse Effects

Since properly designed toxicological studies identify doses at which test substance-related changes occur, decisions must be made on which observed changes are considered adverse effects and that also are relevant and representative of human response. Repeated exposure to GenX in rodents is associated with peroxisome proliferator-activated receptor alpha (PPARα) effects, but the lack of human relevance of this pathway (PPAR α -dependent toxicity) is very well established (Corton et al. 2017; Felter et al. 2018). Thus, the endpoints of reduced serum lipids, hepatocyte hypertrophy, increased liver weights, and liver tumors in the rodent studies of GenX (Table 1) are not relevant to humans. A strong argument also can be made to treat single-cell necrosis as a non-adverse effect because 1) the effects were minimal-to-mild, 2) were not consistently seen at higher doses or at the lower doses at longer time points, and 3) are therefore likely a result of the adaptive and reversible hypertrophy that is known to be PPAR α -dependent and thus not relevant to human health. As the cell enlarges due to the PPAR α -dependent hypertrophy, minimal-to-mild single-cell necrosis would be expected. However, a non-PPARαdependent pathway for single-cell necrosis cannot be ruled out at this time and single-cell necrosis is usually considered adverse when correlative enzyme activation is observed (Thoolen et al. 2010; Maronpot et al. 2010; Hall et al. 2012.) Thus, to be conservative, one could consider the minimal-to-mild single-cell necrosis as an adverse effect in the subchronic/subacute studies.

Use of NOAEL versus Benchmark Dose (BMD) to set Point of Departure (POD)

The US EPA risk assessment process includes an analysis of the dose-response relationship between exposure and adverse health-related outcomes and follows a two-step process: (1) defining a point of departure (POD) and (2) extrapolating from the POD for relevance to human exposure. The NOAEL has been used as the POD for many years, but recognizing the limitations of this approach, the US EPA has adopted an alternative approach called the benchmark dose (BMD) method (US EPA 2012). The BMD method involves statistical modeling of dose-response data and is particularly helpful at incorporating data from multiple related studies and extrapolations near the low end of the exposure range. The BMD analysis results in a BMDL (benchmark dose lower confidence level) that is used as the POD instead of the NOAEL. Not all data sets are amenable to BMD modeling and different models within the BMD software can provide slightly different results. BMD modeling was performed on the data available for GenX using the latest US EPA BMD software (US EPA 2017). A brief summary of this analysis is provided below. BMD analysis was performed on males only because in all studies males were more sensitive than females and would provide the lowest BMDL and POD. A benchmark response (BMR) of 10% extra risk was applied to all dichotomous model types.

The 2-year chronic rat study data (Craig 2013) were not amenable to BMD modeling owing to the lack of observed effects at the lower doses and a wide range between the LOAEL (50 mg/k/day) and NOAEL (1 mg/kg/day), resulting in a best-fit BMDL of 38.2 mg/kg/day (for liver centrilobular necrosis) which is much higher than the NOAEL. Thus, the appropriate POD for the 2-year chronic study is the more conservative NOAEL of 1 mg/kg/day.

For the subchronic mouse studies, the data for male mice were combined for the 90-day subchronic (MacKenzie 2010) and 70+ day DART (Edwards 2010b) studies because the exposure durations were similar, the mice were the same strain (Crl:CD1(ICR)) and single-cell necrosis in the liver was the common adverse but reversible endpoint reported by both studies. These data were amenable to BMD analysis and resulted in a best-fit BMDL of 0.23 mg/kg/day using the multistage model. This model had the second highest p-value (0.9944) and the lowest Akaike's Information Criterion, AIC, value (50.51). The multistage model also resulted in the second lowest BMDL value, where the lowest BMDL value resulted in a very poor fit (p-value 0.0592 and AIC Value 58.40). The multistage model also is the most conservative BMDL that has an acceptable statistical fit (BMDL range was 0.228 – 0.339 mg/kg/day). Although the chronic rat data are the most appropriate and preferred data for setting the POD, the 0.23 mg/kg/day BMDL has a stronger scientific basis for use as the POD than the 0.1 mg/kg/day NOAEL from the subchronic/subacute studies and is used below for comparative purposes.

Uncertainty Factors (UF)

A subchronic-to-chronic UF would not be needed if the 2-year chronic rat study is used to derive the POD. However, if the subchronic study data were used then consideration must be given to the value of this UF. The US EPA states "A default value of 10 for this UF is applied to the NOAEL/LOAEL or BMDL/BMCL from the subchronic study on the assumption that effects from a

given compound in a subchronic study occur at a 10-fold higher concentration than in a corresponding (but absent) chronic study" (US EPA 2014b). This condition is not true for GenX, since a complete 2-year chronic rat study is available for GenX (Craig 2013). Moreover, the rationale for using a subchronic-to-chronic UF = 10 is to account for possible effects "at a 10-fold higher concentration" in a chronic study (US EPA 2014b). However, in the case of GenX the opposite is true. In the mouse studies, there is no single-cell necrosis progression or higher NOAEL/LOAEL with the longer exposure duration study (90 days) compared to the shorter duration studies (28 days and 70+ days). Thus, the subchronic data do not support the use of a default subchronic-to-chronic UF = 10 or even a UF = 3. Rather, the data support a UF = 1 given that longer duration and higher dose did not demonstrate increased severity or incidence of any adverse endpoints.

The US EPA uses a default interspecies UF = 10 under the assumption that humans may be more susceptible/sensitive to a chemical than the rat or mouse used in the toxicity study, but the US EPA also allows for this 10-fold UF to be reduced if there is evidence that that humans are less sensitive (US EPA 2014c). Although definitive human data are lacking, there is good reason to assume that humans are less sensitive than rats and mice to the reported GenX toxicities (single-cell necrosis in subchronic studies and centrilobular necrosis in chronic study) due to the very strong **PPAR\alpha-dependent** toxicities that are not relevant in humans. This would justify setting the interspecies UF = 3.

Relative Source Contribution

The health advisory may include a provision to account for other possible sources of GenX besides drinking water (such as food, inhalation, and dermal absorption) by using a relative source contribution (RSC) to apportion exposure to different sources. Given the chemical properties of the environmentally-relevant form of GenX (e.g., highly water soluble) one would not expect significant amounts of GenX in the food of the general population. This is consistent with a recent study in the Netherlands where GenX was measured in vegetable crops grown in private gardens in the vicinity of the Dordrecht Chemours site where GenX is known to have been released to the environment (Mengelers 2018). The study concluded 1) that very low concentrations of GenX were found in some samples very near the site, but these levels did not exceed exposure thresholds and 2) that beyond 1 km distance from the site, consumption of vegetables did not contribute significantly to GenX exposure. Also, as pointed out by the Centers for Disease Control (CDC), GenX is unlikely to be absorbed through the skin (Moore 2017). Finally, due to its high water solubility and low volatility, any GenX emitted to air would return to the ground though wet deposition. This, combined with the very low volatility of GenX, means that inhalation would not contribute significantly to GenX exposure for the general population. Thus, assuming that 100% or nearly 100% of GenX exposure is through drinking water is probably accurate for most people. However, since we do not yet have enough data on all relevant exposure pathways, for the purpose of this paper the more protective assumption that only 20% of GenX exposure is through drinking water will be used until additional data become available. This would set the RSC value for water to be 20% (RSC = 0.2) in the risk equation.

Derivation of Proposed Health Advisory Value for GenX

Using the above discussion as a basis for deriving a health advisory for GenX, three possible values are listed in Table 2 along with the parameters and assumptions used to derive the value. The values listed in green columns are based on the 2-year chronic study in rat and thus are scientifically defensible in matching the lifetime exposure health advisory to the lifetime chronic study in the rat. The value listed in the yellow column is provided for comparative purposes only to show the value one would derive in the absence of the chronic study data (using the BMD method with subchronic data from the mouse as described above). Based on these analyses, a conservative lifetime health advisory value would be 70,000 ng/L for GenX.

Table 2. Alternative health advisory values based on available data, with assumptions listed.

	US EPA Benchmark Dose Method (subchronic)	Lifetime Exposure Assessment (chronic)	Lifetime Exposure Assessment Adjustment 1 (chronic)
Health Advisory (ng/L or ppt)	16,100	70,000	233,300
Population Assumed	Adult Lifetime Exposure	Adult Lifetime Exposure	Adult Lifetime Exposure
Key Study Used	90-day and 70+ day subchronic mouse	2-year chronic rat	2-year chronic rat
Relative Source Contribution for Water	0.2	0.2	0,2
Subchronic-Chronic UF	1	Not applicable	Not applicable
Interspecies UF	10	10	3
Intraspecies UF	10	10	10

All values are based on lifetime exposure to an adult using 70 kg bodyweight and 2.0 L/day drinking water intake that is the standard in many countries and many states within the US. The US EPA has recently updated bodyweight to be 80 kg and drinking water intake to be either 2.4 L/day or 2.5 L/day (US EPA 2015). The values in Table 2 can be adjusted accordingly depending on local standards/rules.

RISK CHARACTERIZATION

Toxicokinetics and Elimination Half-lives

The toxicokinetics of GenX was initially studied in male and female rats that were given either a 10 mg/kg or a 30 mg/kg dose (Gannon 2008). Concentrations of GenX were followed in the plasma of the rats at 14 time points up to 168 hours (Fig. 2) and GenX was measured in the liver and adipose tissues at time of sacrifice (168 hours). Clearance times, defined as the time for 98.4% of the initial GenX concentration in the plasma to be cleared, were 28 hours (10 mg/kg dose) and 22 hours (30 mg/kg dose) for males and 8 hours (10 mg/kg does) and 4 hours (30 mg/kg dose) for females. At 168 hours, the ratio of GenX in liver:plasma for males was 0.64 (10 mg/kg dose) and 0.71 (30 mg/kg dose). The liver:plasma GenX ratio could not be calculated for females because GenX was not detectable in plasma in females after 120 hours and was never detected

in the liver of females at either dose. GenX was never detected in adipose tissue in either males or females at either dose, indicating that GenX is not stored in the fat tissue. Overall, these data show very fast clearance of GenX from rats.

A follow-on study of the absorption, distribution, metabolism, and elimination (ADME) and kinetic behavior of GenX was performed in rats, mice, and cynomolgus monkeys (Gannon et al. 2016). A single dose of GenX was given by oral and intravenous routes of exposure and followed in both plasma and urine. GenX was rapidly and completely absorbed in both rats and mice and then rapidly eliminated in the urine with elimination half-lives of approximately 5 hours in rats and 20 hours in mice. In monkeys, rats, and mice GenX undergoes a rapid, biphasic elimination with a very fast alpha phase and a slower beta phase. The beta phase indicated no potential for accumulation after multiple dosing in rats or monkeys (Gannon et al. 2016), consistent with the earlier study of rats showing no detectable GenX in adipose tissue (Gannon 2008). Both *in vivo* and *in vitro* experiments demonstrated that GenX is not metabolized. This comparative pharmacokinetic study in rats, mice, and monkeys indicated that the rat, and not mice, is more similar to the monkey and the authors concluded that the rat is therefore a more appropriate rodent model than that of the mouse for a pharmacokinetics assessment of GenX behavior in primates (Gannon et al. 2016.) This provides further justification for using the 2-year chronic rat study rather than the short-term mouse studies (Table 1).

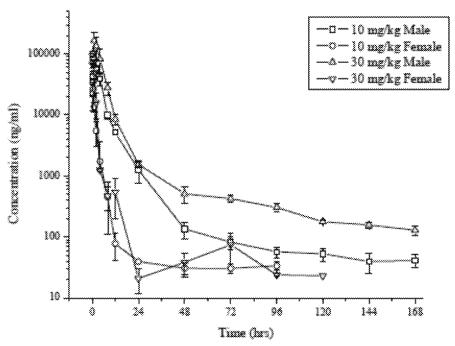


Figure 2. Toxicokinetics of GenX in plasma of male and female rats at two doses (from Gannon 2008). Clearance times (time for 98.4% of initial value to be cleared from the plasma) where 28 hours (10 mg/kg dose) and 22 hours (30 mg/kg dose) for males and 8 hours (10 mg/kg does) and 4 hours (30 mg/kg dose) for females. The ratio of GenX in liver:plasma for males at time of sacrifice (168 hours) was 0.64 (10 mg/kg dose) and 0.71 (30 mg/kg dose). GenX was not detectable in female plasma after 120 hours and was never detected in the liver of females at either dose. GenX was never detected in fat tissue in either males or females at any time. All data are from Gannon 2008.

No estimates for human elimination rates for GenX have been published yet, but values can be estimated from monkey:human elimination rate ratios of other perfluorinated chemicals: PFOA, PFOS and shorter chain fluoropolymers perfluorobutane sulfonate (PFBS) and perfluorobutanoic acid (PFBA). It is likely that the monkey:human elimination rate ratio of GenX would fall between the values of the longer chain PFAS chemicals (PFOA: 66 or 40; PFOS: 16) and the shorter chain PFAS chemicals (PFBS: 7.5; PFBA: 1.9). Using the data from Gannon et al. (2016) for GenX in monkey and a conservative monkey:human ratio of 16 (PFOS) comparison of GenX is made to the legacy compounds PFOA, PFOS and other chemicals regarding elimination rates in humans (Figure 3) using data for humans for the other chemicals (US EPA 2016a, US EPA 2016b, Gannon et al. 2016, Olsen et al. 2007, Bartell et al. 2010, Wang et al 2013.). Clearly GenX has a much faster estimated elimination rate in humans compared to PFOA, PFOS and other legacy chemicals with a half-life range of 4 hours to 6 days depending on which monkey:human elimination rate ratio is used as a surrogate for GenX. The elimination curve for GenX shown in Figure 3 corresponds to a half-life in humans of 1.8 days and is based on the monkey:human elimination rate ratio of 16 for PFOS.

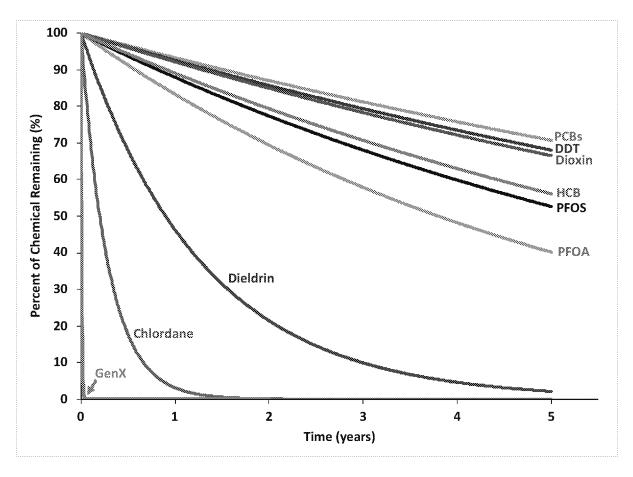


Figure 3. Estimated elimination curves in humans showing percent of various chemicals remaining in the body as a function of time. The elimination curve for GenX is based on a conservative assumption that GenX follows the same monkey:human half-life ratio as PFOS (16). Using the monkey:human ratio for the shorter-chain PFBS (7.5) or PFBA (1.9) would lead to even more rapid loss of GenX in humans. See text for data sources and explanation of estimation method.

Margin of Safety or Exposure: Comparing Measured Water Values of GenX to the Proposed Health Advisory

Recent data on GenX in drinking water from North Carolina, USA are used to provide some perspective on how measured values of GenX compare to the proposed health advisory. A study measuring GenX in tap water from 198 homes potentially affected in NC reported GenX values ranging from non-detectable to nearly 100 ng/L with an average of 45 ng/L (Kotlarz 2018). The highest value measured represents a margin of exposure of greater than 700 compared to the proposed health advisory of 70,000 ng/L. Similar margins of exposure are found for data in the Cape Fear River, NC, USA and in homeowner wells near the source of GenX in that area.

CONCLUSIONS

There is sufficient information to perform a preliminary analysis to provide guidance to scientists, regulators and the public on the potential for adverse health effects associated with exposure to GenX through drinking water. A 2-year chronic study with rats provides the best available data on GenX toxicity for deriving a lifetime exposure health advisory. The NOAEL for this study was 1.0 mg/kg/day and was used as the point of departure for deriving the health advisory. Using protective uncertainty factors of 10 for both interspecies and intraspecies and a conservative relative source contribution from water of 20%, a conservative lifetime health advisory value would be 70,000 ng/L for GenX. Data on the elimination of GenX from mammals indicate its half-life in humans is in the range of 4 hours to 6 days, and most likely at the low end of that range, indicating no potential for significant accumulation in humans. Using data from North Carolina, USA where GenX has been detected in drinking water and drinking water sources, indicates a margin of safety (exposure) in excess of 700 in the most recent tap water sampling. In other words, for a consumer to approach the derived health threshold, they would have to consume 1,400 L of water per day from the most contaminated tap for their entire life.

REFERENCES

Bartell SM, Calafat AM, Lyu C, Kato K, Ryan PB, Steenland K. 2010. Rate of Decline in Serum PFOA Concentrations after Granular Activated Carbon Filtration at Two Public Water Systems in Ohio and West Virginia. Environ Health Perspect 118: 222-228.

Beekman M, Zweers P, Muller A, de Vries W, Janssen P, Zeilmaker M. 2016. Evaluation of substances used in the GenX technology by Chemours, Dordrecht. RIVM Letter report 2016-0174, 92pp.

Bloxham PA. 2008. Estimation of the Adsorption Coefficient (K_{oc}) of HFPO Dimer Acid Ammonium Salt on Soil and Sludge. Report by Dupont, Study No. DuPont-17568-1675. 11 pp.

- Buck RC. 2015. Toxicology data for alternative "Short-Chain" fluorinated substances. In: DeWitt, J.C. (Ed.), Toxicological Effects of Perfluoroalkyl and Polyfluoroalkyl Substances. Humana Press, New York, pp 451-477.
- Buck RC, Franklin J, Berger U, Conder JM, Cousins IT, de Voogt P, Jensen AA, Kannan K, Mabury SA, van Leeuwen SPJ. 2011. Perfluoroalkyl and polyfluoroalkyl substances in the environment: terminology, classification, and origins. Integr. Environ. Assess. Manag. 7, 513–541.
- Caverly Rae JM, Craig L, Slone TW, Frame SR, Buxton LW, Kennedy GL. 2015. Evaluation of chronic toxicity and carcinogenicity of ammonium 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoate in Sprague-Dawley rats. Toxicol. Rep. 2, 939-949.
- Corton C, Peters JM, Klauning JE. 2017. The PPARα-dependent rodent liver tumor response is not relevant to humans: addressing misconceptions. Archives of Toxicology. https://doi.org/10.1007/s00204-017-2094-7.
- Craig L. 2013. Combined Chronic Toxicity/Oncogenicity Study 2-Year Oral Gavage Study in Rats. Report by MPI Research Inc. to E.I du Pont de Nemours and Company. Dupont 18405-1238. 4037 pp.
- ECHA 2017a. ammonium 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)propanoate, Bioaccumulation: aquatic/sediment. Accessed 24 April 2018 at https://echa.europa.eu/registration-dossier/-/registered-dossier/2679/5/4/2.
- ECHA 2017b. ammonium 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)propanoate, Bioaccumulation: terrestrial. Accessed 24 April 2018 at https://echa.europa.eu/registration-dossier/-/registered-dossier/2679/5/4/3.
- Edwards T. 2010a. An Oral (Gavage) Prenatal Developmental Toxicity Study of H-28547 in Rats. Report by WIL Research Laboratories to E.I du Pont de Nemours and Company. Dupont 18405-841. 388 pp.
- Edwards T. 2010b. An Oral (Gavage) Reproduction/Developmental Toxicity Screening Study of H-25848 in Mice. Report by WIL Research Laboratories to E.I du Pont de Nemours and Company. Dupont 18405-1037, 2072 pp.
- Feiring AE. 1994. Fluoroplastics. In Organofluorine Chemistry. Principles and Commercial Applications. Plenum Press, pp 339-364.
- Felter SP, Foreman JE, Boobis A, Corton JC, Doi AM, Flowers L, Goodman J, Haber LT, Jacobs A, Klaunig JE, Lynch AM, Moggs J, Pandiri A. 2018. Human relevance of rodent liver tumors: Key insights from a Toxicology Forum workshop on nongenotoxic modes of action. Regul Toxicol Pharmacol 92:1–7.

- Gannon SA. 2008. Biopersistence and Pharmacokinetic Screen in the Rat. Report by Critical path Services to E.I du Pont de Nemours and Company. Dupont 24286. 8 pp.
- Gannon SA, Fasano WJ, Mawn MP, Nabb DL, Buck RC, Buxton LW, Jepson GW, Frame SR. 2016. Absorption, distribution, metabolism and excretion of 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)propionic acid ammonium salt in rat, mouse, and cynomolgus monkey. Toxicology 340, 1-9.
- Gebbink WA, van Asseldonk L, van Leeuwen S. 2017. Presence of emerging per- and polyfluoroalkyl substances (PFASs) in river and drinking water near a fluorochemical production plant in the Netherlands. Environ. Sci. Technol. 2017, 51, 11057–11065.
- Gordon, S.C. 2011. Toxicological evaluation of ammonium 4,8-dioxa-3H-perfluorononanoate, a new emulsifier to replace ammonium perfluorooctanoate in fluoropolymer manufacturing. Reg. Toxicol. Pharm. 59, 64e80.
- Haas MC. 2008a. An Oral (Gavage) Toxicity Study of H-28397 in Rats with a 28-day Recovery. Report by WIL Research Laboratories to E.I du Pont de Nemours and Company. Dupont 24447. 1028 pp.
- Haas MC. 2008b. An Oral (Gavage) Toxicity Study of H-28397 in Mice with a 28-day Recovery. Report by WIL Research Laboratories to E.I du Pont de Nemours and Company. Dupont 24459. 1000 pp.
- Haas MC. 2009. A 90-Day Oral (Gavage) Study of H-28548 in Rats with a 28-Day Recovery. Report by WIL Research Laboratories to E.I du Pont de Nemours and Company. Dupont 17751-1026. 2076 pp.
- Hall AP et al. 2012. Liver hypertrophy: a review of adaptive (adverse and non-adverse) changes conclusions from the 3rd International ESTP Expert Workshop. Toxicologic Pathology. 40:971–994.
- Heydebreck F, Tang J, Xie Z, Ebinghaus R. 2015. Alternative and Legacy Perfluoroalkyl Substances: Differences between European and Chinese River/Estuary Systems. Environ. Sci. Technol. 2015, 49 (14), 8386–8395.
- Hoke RA, Ferrell BD, Sloman TL, Buck RC, Buxton LW. 2016. Aquatic hazard, bioaccumulation and screening risk assessment for ammonium 2,3,3,3-tetrafluoro-2-(heptafluoropropoxy)-propanoate. Chemosphere 149:336-342.
- Kissa E. 2001. Fluorinated surfactants and repellents, vol 97, Surfactant science series. Marcel Dekker, NY, New York.

- Kotlarz N. 2018. GenX Exposure Study. Presentation made at UNC Wilmington 17 April 2018.
- MacKenzie SA. 2010. H-28548: Subchronic Toxicity 90-Day Gavage Study in Mice. Report by E.I du Pont de Nemours and Company. Dupont 18405-1307. 339 pp.
- Maronpot RR et al. 2010. Hepatic enzyme induction: Histopathology. Toxicologic Pathology. 38: 776–95.
- Mengelers MJB, te Biesebeek JD, Schipper M, Slob W, Boon PE. 2018. Risk assessment of GenX and PFOA in vegetable garden crops in Dordrecht, Papendrecht and Sliedrecht. RIVM Letter report 2018-0017, 68pp.
- Mitsubishi Chemical. 2009. Ready Biodegradability Test of FRD903. Report by Mitsubishi Chemical to du Pont-Mitsui Fluorochemicals Company, Ltd. 30 pp.
- Moore Z. 2017. Presentation by Dr. Zack Moore, NC State Epidemiologist to the Secretaries' Science Advisory Board (SAB) on 4 December 2017. Accessed 25 April 2018 https://files.nc.gov/ncdeq/GenX/SAB/GenX%20Health%20Studies%20and%20Advisories%2 0SSAB%2012_4_2017.pdf
- Nabb D. 2008a. Repeated Dose Oral Toxicity 7-Day Gavage Study in Mice [HFPO Dimer Acid]. Memo Report by Dupont, Report No. DuPont-25281, 11pp.
- Nabb D. 2008b. Repeated Dose Oral Toxicity 7-Day Gavage Study in Mice [HFPO Dimer Acid Ammonium Salt]. Memo Report by Dupont, Report No. DuPont-24010, 11pp.
- NC DEQ. 2018. GenX Investigation. Retrieved from https://deq.nc.gov/news/hot-topics/genx-investigation. Accessed 26 April 2018.
- Olsen GW, Burris JM, Ehresman DJ, Froehlich JW, Seacat AM, Butenhoff JL, Zobel LR. 2007. Half-life of serum elimination of perfluorooctanesulfonate, perfluorohexanesulfonate, and perfluorooctanoate in retired fluorochemical production workers. Environ Health Perspect 115:1298–1305.
- Ritter SK. 2010. Fluorochemicals go short. Chem. Eng. News 88, 12-17.
- Rushing BR, Hu Q, Franklin JN, McMahen RL, Dagnino S, Higgins CP, Strynar MJ, DeWitt JC. 2017. Evaluation of the Immunomodulatory Effects of 2,3,3,3- Tetrafluoro-2- (Heptafluoropropoxy)-Propanoate in C57BL/6 Mice. Tox Sci., 156(1), 2017, 179–189.
- Sheng N, Cui R, Wang J, Guo Y, Wang J, Dai J. 2018. Cytotoxicity of novel fluorinated alternatives to long-chain perfluoroalkyl substances to human liver cell line and their binding capacity to human liver fatty acid binding protein. Arch Toxicol 92:359–369.

- Sun M, Arevalo E, Strynar M, Lindstrom A, Richardson M, Kearns B, et al. 2016. Legacy and emerging perfluoroalkyl substances are important drinking water contaminants in the Cape Fear River Watershed of North Carolina. Environ Sci Technol Letters, 3:415–419.
- Thoolen B et al. 2010. Proliferative and Nonproliferative Lesions of the Rat and Mouse Hepatobiliary System, Toxicologic Pathology, 38: 5S-81S.
- UNCW (anonymous). 2018. Report to the Environmental Review Commission from The University of North Carolina at Wilmington Regarding the Implementation of Section 20.(a)(2) of House Bill 56 (S.L. 2017-209) 18pp. Retrieved (April 25 2018) from https://www.ncleg.net/documentsites/committees/ERC/ERC%20Reports%20Received/201 8/CFPUA%20and%20UNC-W/2018-April%20HB%2056%20UNCW%20Rpt.pdf
- US EPA. 2012. Benchmark Dose Technical Guidance. EPA/100/R-12/001 June 2012, 99 pp. Accessed 25 April 2018 https://www.epa.gov/risk/benchmark-dose-technical-guidance
- US EPA. 2014a. A Review of the Reference Dose and Reference Concentration Process. EPA/630/P-02/002F December 2002 Final Report pp 3-9. Accessed 25 April 2018 https://www.epa.gov/sites/production/files/2014-12/documents/rfd-final.pdf
- US EPA. 2014b. A Review of the Reference Dose and Reference Concentration Process. EPA/630/P-02/002F December 2002 Final Report pp 4-45. Accessed 25 April 2018 https://www.epa.gov/sites/production/files/2014-12/documents/rfd-final.pdf
- US EPA. 2014c. Guidance for Applying Quantitative Data to Develop Data-Derived Extrapolation Factors for Interspecies and Intraspecies Extrapolation, EPA/100/R-14/002F September 2014 Accessed 25 April 2018 https://www.epa.gov/sites/production/files/2015-01/documents/ddef-final.pdf
- US EPA. 2015. EPA Response to Scientific Views from the Public on Draft Updated National Recommended Water Quality Criteria for the Protection of Human Health (Docket ID No. EPA-HQ-OW-2014-0135). EPA 822-R-15-001, 48 pp.
- US EPA. 2016a. Drinking Water Health Advisory for Perfluorooctanoic Acid (PFOA). EPA Document Number: 822-R-16-005 May 2016 103pp. https://www.epa.gov/sites/production/files/2016-05/documents/pfoa_health_advisory_final_508.pdf
- US EPA. 2016b. Drinking Water Health Advisory for Perfluorooctane Sulfonate (PFOS). EPA Document Number: 822-R-16-004 May 2016 88pp. https://www.epa.gov/sites/production/files/2016-05/documents/pfos health advisory final 508.pdf
- US EPA. 2017. Benchmark Dose Software, BMDS 2.7 (rel. 2017-08-18) Accessed 25 April 2018 https://www.epa.gov/bmds/download-benchmark-dose-software-bmds

Wang Z, Cousins IT, Scheringer M, Hungerbühler K. 2013. Fluorinated alternatives to long-chain perfluoroalkyl carboxylic acids (PFCAs), perfluoroalkane sulfonic acids (PFSAs) and their potential precursors. Environ. Int. 60;242–248.

Table 1. Summary of Rodent Toxicity Studies. Observations and effects are taken directly from statements of the study authors in the toxicology reports. All studies were conducted under GLP.

Study Reference Regulatory Guideline Species/Study Type Exposure/Duration	Dose (mg/kg/day)	Effects and Observations in Males	Effects and Observations in Females
Key Study Identific	ed by Europea	n Chemicals Agency and used in this Study to De	erive the Drinking Water Health Advisory Value
Craig 2013. Combined	0.1	No adverse effects were observed at this dose	Not applicable, dose not given
Chronic Toxicity/Oncogenicity	1	NOAEL: no adverse effects were observed at this dose	No adverse effects were observed at this dose
Study 2-Year Oral Gavage Study in Rats. (ECHA Study 001) Data also summarized in Caverly Rae et al. 2015	50	LOAEL: Increases in focal cystic degeneration, focal necrosis, and centrilobular necrosis of the liver, with associated increases in cytotoxic liver enzymes, and equivocal increases in pancreatic acinar cell tumours and testicular interstitial (Leydig) cell tumours	NOAEL: No effects were observed at this dose
OECD 453 (GLP) Rat/Chronic-Cancer Gavage/2-year	500	Not applicable, dose not given	LOAEL: Reductions in body weight, body weight gain, and food efficiency; mild decreases in red cell mass; increases in individual cell necrosis in the liver, hyperplasia and/or inflammation in the nonglandular stomach and tongue; a increase in incidence and severity of microscopic pathology in the kidneys; and an increase in hepatocellular adenomas and carcinomas,
Supporting Study Ide	ntified by the I	European Chemicals Agency and <u>not used</u> to De	rive the Drinking Water Health Advisory Value
Haas 2009. A 90-Day	0.1	No adverse effects were observed at this dose	Not applicable, dose not given
Oral (Gavage) Study of H-28548 in Rats with a 28-Day Recovery (ECHA Study 002) OECD 408 (GLP)	10	NOAEL: Study authors state that clinical chemistry changes were associated with PPARα activation (decreased cholesterol, decreased globulin, increased albumin, increased A/G ratio); increased liver weights, hepatocellular hypertrophy (reversible); increased kidney weights (not adverse)	No adverse effects were observed at this dose; increased kidney weights (not adverse)
Rat/Subchronic	100	LOAEL: Erythrocyte changes associated with regenerative anemia (reversible)	NOAEL: Study authors noted that the clinical chemistry changes (decreased cholesterol and globulin) were associated with PPAR-alpha activation

Study Reference Regulatory Guideline Species/Study Type Exposure/Duration	Dose (mg/kg/day)	Effects and Observations in Males	Effects and Observations in Females
Gavage/90-day 4-wk recovery	1000	Not applicable, dose not given	LOAEL: Three mortalities (two were associated with treatment); erythrocyte changes associated with regenerative anemia (reversible); additional clinical chemistry changes (lower total protein, bilirubin, and GGT levels; higher A/G ratio); increased liver weights, hepatocellular hypertrophy (reversible)
Supporting Study Id	T	e European Chemicals Agency and <u>not used</u> to Detail to	
	0.3	Decreased cholesterol; increased hepatic peroxisomal beta-oxidation (considered reversible)	Not applicable, dose not given
Haas 2008a. An Oral (Gavage) Toxicity Study of H-28397 in Rats with a 28-day Recovery (ECHA Study 004) OECD 407 (GLP)	3	Study authors state that clinical chemistry changes were associated with PPARα (decreased globulin, increased albumin, increased A/G ratio, decreased triglycerides (not significant at other doses)); minimal decreases in red cell mass parameters; increased liver weights and kidney weights; multifocal centrilobular liver hypertrophy (all effects were considered reversible and non-adverse by the study authors)	No adverse effects were observed at this dose
Rat/Subacute	30	NOAEL : increased hepatic microsomal cytochrome P- 450 enzyme (considered reversible and non-adverse)	Increased hepatic peroxisomal beta-oxidation (considered reversible and non-adverse)
Gavage/28-day 4-wk recovery	300	Not applicable, dose not given	NOAEL: Study authors reported that clinical chemistry changes were associated with PPARα activation (decreased globulin, increased albumin, and increased A/G ratio); increased relative liver weights; multifocal centrilobular hypertrophy (all effects reversible and nonadverse)
Supporting Study Id	dentified by the	e European Chemicals Agency and <u>not used</u> to De	erive the Drinking Water Health Advisory Value
Edwards 2010a. An Oral (Gavage) Prenatal Developmental	10	Not applicable, dose not given	MATERNAL (F0) / FETAL (F1 both genders) F0: Maternal NOAEL; no adverse effects were observed a this dose

Study Reference Regulatory Guideline Species/Study Type Exposure/Duration	Dose (mg/kg/day)	Effects and Observations in Males	Effects and Observations in Females
Toxicity Study of H- 28547 in Rats			F1: Developmental NOAEL ; no adverse effects were observed at this dose
OECD 414 (GLP)	100	Not applicable, dose not given	F0: LOAEL : Early deliveries; lower gravid uterine weights; higher liver weights, focal necrosis of liver F1: Reduced mean fetal weights
Rat/Developmental Drinking water/ Gestation Day 6-20	1000	Not applicable, dose not given	F0: One mortality; reduced body weight and body weight gains; edematous pancreas (2 females); higher kidney weights, hepatocellular hypertrophy F1: Higher mean litter skeletal variations
Supporting Study Id	dentified by the	e European Chemicals Agency and <u>not used</u> to D	erive the Drinking Water Health Advisory Value
Haas 2008b. An Oral (Gavage) Toxicity Study of H-28397 in Mice with a 28-day Recovery	0.1	NOAEL: increased hepatic peroxisomal beta-oxidation was found at all doses, however study authors indicate this observation was completely reversible and consistent with PPARα agonist and considered non-adverse.	No adverse effects were observed at this dose
(ECHA Study 005) OECD 407 (GLP) Mouse/Subacute	3	Non-adverse effects were PPARα agonist related (increased beta-oxidation in the liver, increased liver weights, hepatocellular hypertrophy, and changes in serum lipids and proteins) or reversible (increased body weights, minimal decreases in red cell mass parameters and increased adrenal weights and adrenal cortical	NOAEL: Decreased globulin and A/G ratio (reversible); increased liver weights (reversible at 3 mg/kg/day, but not 30 mg/kg/day); changes in liver enzymes, increased hepatic peroxisomal beta-oxidation at doses ≥ 3 mg/kg/day (reversible). All observations considered non-adverse.
Gavage/28-day 4-wk recovery		hypertrophy. Adverse effects were single cell necrosis of hepatocytes (4/10; minimal) and correlative increases in hepatic microsomal cytochrome P-450 enzyme content liver enzymes, but both of these also were reversible.	

Study Reference Regulatory Guideline Species/Study Type Exposure/Duration	Dose (mg/kg/day)	Effects and Observations in Males	Effects and Observations in Females
	30	Increased body weights; increased monocytes and large unstained cells (not associated with other hematological effects), increased albumin (reversible); liver enzymes increased (ALT, AST, SDH, ALKP; all were reversible except SDH), multifocal single cell hepatocellular necrosis (10/10; minimal, reversible), decreased chloride and increased BUN (not associated with correlative microscopic changes); adrenal cortical hypertrophy (reversible); increased mitoses distributed multifocally throughout the liver (reversible)	Liver enzymes increased (ALKP and SDH; reversible); decreased uterine weights (reversible); reversible liver changes: hepatocellular hypertrophy, increased mitoses distributed multifocally throughout the liver, multifocal single cell hepatocellular necrosis; increased number of animals in diestrus stage (likely secondary to systemic stress indicated by adrenal hypertrophy; not observed in recovery group)
Supporting Study Id	dentified by the	European Chemicals Agency and <u>not used</u> to D	erive the Drinking Water Health Advisory Value
MacKenzie 2010. H-	0.1	No adverse effects were observed at this dose	Increased monocytes (not observed at higher doses)
28548: Subchronic Toxicity 90-Day Gavage	0.5	NOAEL: increased liver weights, hepatocellular hypertrophy (reversible and PPARα dependent)	NOAEL: No adverse effects were observed at this dose
Study in Mice (ECHA Study 003) OECD 408 (GLP) Mouse/Subchronic Gavage/90-day	5	LOAEL: Increased total bile acids and liver enzymes along with increased liver weights, and microscopic changes (increased hypertrophy, single cell necrosis (10/10, minimal), mitotic figures and/or Kupffer cell pigment); increased albumin and total protein, reduced cholesterol, decreased potassium; increased adrenal weights with cortical hypertrophy, increased kidney weight with minimal tubular epithelial hypertrophy, reduced spleen weight (no pathological changes)	LOAEL: Increased total bile acids and liver enzymes along with increased liver weights and microscopic changes (increased hypertrophy, single cell necrosis (1/10, minimal), mitotic figures and/or Kupffer cell pigment); increased albumin, decreased bilirubin, decreased potassium
Supporting Study Id	dentified by the	e European Chemicals Agency and <u>not used</u> to D	erive the Drinking Water Health Advisory Value
Edwards 2010b. An Oral	0.1	NOAEL: No adverse effects were observed at this dose	No adverse effects were observed at this dose
(Gavage) Reproduction/ Developmental Toxicity	0.5	Hepatocellular hypertrophy (minimal/mild) and increased liver weights; 5 of 24 animals with minimal	NOAEL: Hepatocellular hypertrophy (minimal/mild) and increased liver weights

Study Reference Regulatory Guideline Species/Study Type Exposure/Duration	Dose (mg/kg/day)	Effects and Observations in Males	Effects and Observations in Females
Screening Study of H- 25848 in Mice		single cell necrosis (versus 1 of 24 in controls) was considered to be test-substance related	
OECD 421 (GLP)		Various liver lesions including increases in single cell necrosis (24/24 animals 4 minimal, 17 mild and 3 moderate), mitotic figures, and lipofuscin pigment	liver lesions including increases in single cell necrosis (21/24 animals) and focal necrosis, mitotic figures, and lipofuscin pigment
Mouse Developmental/			
reproductive toxicity	5		NOAEL for reproductive toxicity : There were no effects on reproduction (mating, fertility, or copulation indices,
Gavage/			number of days between pairing and coitus, and gestation
F0 males: 70+ days			length) at any dose.
F0 females: 57+ days			

NOAEL: No Observed Adverse Effect Level; LOAEL: Lowest Observed Adverse Effect Level; A/G: albumin to globulin; PPARα: Peroxisome proliferator-activated receptor-alpha; RBC: red blood cell; BUN: blood urea nitrogen